

## Indicanine A, a New 3-Phenylcoumarin from Root Bark of *Erythrina indica*<sup>1</sup>

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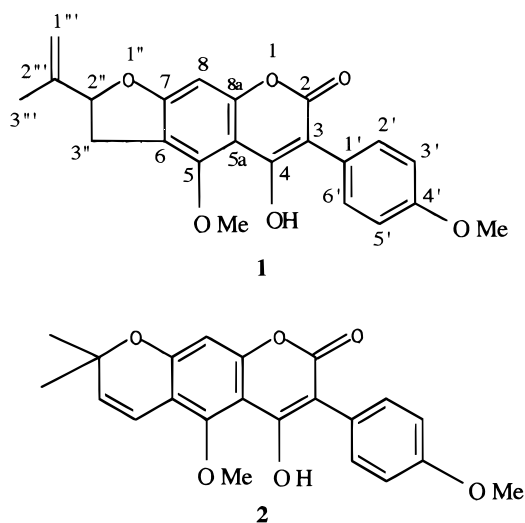
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A new 3-phenylcoumarin, indicanine A (**1**), has been isolated from the root bark of the African medicinal plant *Erythrina indica*, together with three known compounds, robustic acid (**2**), daidzein, and 8-prenyldaidzein. The structure of the new compound was characterized, as 4-hydroxy-5-methoxy-3-(4'-methoxyphenyl)-2''-(1-methylethenyl)dihydrofurano[4'',5'':6,7]coumarin by means of extensive spectroscopic analyses. The compounds were found to be devoid of *in vitro* antibacterial activity.

Plants species belonging to genus *Erythrina* (Papilionaceae) are well-known for elaborating, from seeds and leaves, alkaloids possessing cardiovascular effects<sup>2</sup> and, from stem and root barks, isoflavonoids (pterocarpanes, isoflavones, isoflavanones)<sup>3–5</sup> and flavonoids (chalcones, flavanones),<sup>6,7</sup> of which some exhibit antibacterial and antifungal activity,<sup>7,8</sup> as well as inhibited platelet aggregation.<sup>8</sup>

As part of our continuing investigation on the phenolic metabolites from *Erythrina* species found in Cameroon and elsewhere, we have examined the chemical constituents of the root bark of *E. indica*, a plant used extensively in African folk medicine for the treatment of several diseases, including microbial infections.<sup>9</sup> In this paper, we report the isolation and structure elucidation of a new 3-phenylcoumarin, designated indicanine A, along with three known co-occurring compounds, the phenylcoumarin robustic acid (**2**), and the isoflavones daidzein and 8-prenyldaidzein.



The dried and ground root bark of *E. indica* was successively extracted at room temperature with a mixture of CH<sub>2</sub>Cl<sub>2</sub>–MeOH (1:1) and the extract concentrated to dryness. The residue showed antimicrobial activity against

*Staphylococcus aureus* and *Mycobacterium smegmatis* at <1000 µg/mL in the agar dilution-streak assay.<sup>10</sup> This residue, on repeated column chromatographic separations over Si gel, afforded pure compounds, including indicanine A (**1**) and the three known compounds, robustic acid (**2**),<sup>11,12</sup> daidzein,<sup>13</sup> and 8-prenyldaidzein,<sup>14</sup> which were identified by comparison with reported spectroscopic data.

Compound **1**, mp 175–177 °C, was obtained as an optically active pale yellow amorphous solid, with molecular formula C<sub>22</sub>H<sub>20</sub>O<sub>6</sub>, as established by mass spectrometry. The broad-band decoupling <sup>13</sup>C NMR spectrum of **1** showed 20 carbon signals. The analysis of this spectrum with the aid of *J*<sub>Mod</sub> and DEPT techniques unequivocally indicated the presence of three methyl groups, two methylene carbons, and six methine groups. Thus, there were 11 quaternary carbons, all sp<sup>2</sup>. The IR spectrum showed bands attributable to hydroxyl (3267 cm<sup>-1</sup>), conjugated carbonyl (1645 cm<sup>-1</sup>), and benzene ring (1610 and 1520 cm<sup>-1</sup>). The <sup>1</sup>H NMR signal at δ 10.0 ppm, <sup>13</sup>C NMR signal at δ 162.7 ppm (C-2), and UV absorption bands at 218, 270, 282 sh, 291, 305 sh, and 351 nm were all typical of a 3-phenylcoumarin skeleton.<sup>15,16</sup> These data agreed closely with those reported for robustic acid (**2**).<sup>11,12</sup> Addition of NaOAc did not cause a bathochromic shift, thus suggesting that there is no free phenolic group at C-7. In the <sup>1</sup>H NMR spectrum, the D<sub>2</sub>O exchangeable signal at δ 10.0 ppm is characteristic of 4-OH resonance.<sup>11,12</sup> Also, the <sup>1</sup>H NMR spectrum of **1** shows an AA'BB' spin system of four phenyl protons at δ 7.42 ppm (2H, d, *J* = 8.8 Hz, H-2' and H-6') and δ 6.93 ppm (2H, d, *J* = 8.8 Hz, H-3' and H-5'), indicating the presence of a *para*-substituted ring B and a 1H singlet at δ 6.58 ppm due either to H-6 or H-8 of ring A. Furthermore, the set of signals consisting of two 1H doublets at δ 5.0 ppm (1H, *J* = 1.0 Hz) and δ 4.95 ppm (1H, d, *J* = 1.0 Hz), due to two geminal olefinic protons and a 3H signal at δ 1.77 ppm establish the presence of an isopropenyl substituent. The presence of a dihydrofuran unit was supported by two 3H singlets at δ 4.08 and 3.80 ppm due to two methoxyl groups and two sets of two double doublets at δ 3.58 (1H, dd, *J* = 8.8 and 15.3 Hz) and δ 3.23 ppm (1H, dd, *J* = 7.6 and 15.8 Hz) for the two diastereotopic protons at C-3'' and 1H triplet at δ 5.28 ppm (1H, t, *J* = 8.4 Hz, H-2''). On the basis of these data, compound **1** must be a 4-hydroxy-3-phenyl coumarin substituted in ring A with one methoxyl group at C-5 position and an isopropenyldihydrofuran moiety. This was confirmed by EIMS of indicanine A (**1**), which undergoes RDA fragmen-

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tation via its keto tautomer to give two important ion fragments at  $m/z$  233 and 148. The first ion fragment at  $m/z$  233, clearly showed compound **1** possesses one methoxyl and an isopropenyldihydrofuran moiety on ring A, while the second ion fragment at  $m/z$  148, indicated the second methoxyl group to be located on ring B at C-4' position. Therefore, it remained to be established unambiguously whether the fusion of the isopropenyl dihydrofuran moiety on ring A is linear or angular. This was deduced from NOE difference experiments, which showed no enhancement of the 1H aromatic signal at  $\delta$  6.58 ppm (corresponding either to H-6 or H-8) but enhancement of the signal at  $\delta$  10.0 ppm (4-OH), when the signal at  $\delta$  4.08 ppm (5-OMe) was irradiated. This finding clearly indicated that the single A-ring aromatic proton at  $\delta$  6.58 ppm was located at C-8. Thus, the isopropenyl dihydrofuran unit was fused in a linear manner on ring A. From the above spectroscopic studies, compound **1** was characterized as 4-hydroxy-5-methoxy-3-(4'-hydroxyphenyl)-2''-(1-methylethenyl)dihydrofuran-[4'',5'':6,7]coumarin. This compound, which appears to be novel, has been given the trivial name indicanine A. We were not able to establish the absolute configuration at the C-2'' stereocenter.

All the isolated compounds were tested *in vitro* for their antimicrobial activities against microorganisms, *S. aureus* 209P, *M. smegmatis* ATCC 607, and *Escherichia coli* RL65 using an agar dilution-streak method.<sup>10</sup> None of the compounds showed any significant activity.

## Experimental Section

**General Experimental Procedures.** All melting points were determined on a Kofler hot-stage apparatus and are uncorrected. IR spectra were recorded on a Perkin-Elmer 727B spectrometer in KBr disks. UV spectra were obtained on a Beckman model 25 spectrophotometer. EIMS (ionization voltage, 70 eV) were measured with LKB9000S and Nermag/sidar U 3:1 spectrometers. <sup>1</sup>H and <sup>13</sup>C NMR spectra were recorded on a Varian Gemini 2000 and on a Bruker spectrometers equipped with a 5-mm <sup>1</sup>H and <sup>13</sup>C probe operating at 300 and 75 MHz, respectively, with TMS as internal standard. DEPT and  $J_{\text{Mod}}$  were measured with the usual pulse sequence, and data processing was obtained with standard software.

**Plant Material.** Root bark of *E. indica* was collected in June 1998, at Ibadan, Nigeria. A voucher specimen documenting the collection is on deposit at the National Herbarium, Yaounde, Cameroon.

**Extraction and Isolation.** Air-dried, powdered root bark of *E. indica* (6 kg) was extracted with a mixture of CH<sub>2</sub>Cl<sub>2</sub>-MeOH (1:1) and concentrated to dryness on a rotary evaporator under reduced pressure to afford a viscous mass of CH<sub>2</sub>Cl<sub>2</sub>-MeOH (1:1) extract (200 g). This material was subjected to column chromatography on Si gel (70–230 mesh, ASTM; Merck) packed in *n*-hexane and eluted with *n*-hexane-EtOAc mixture. In all, 200 fractions (ca. 250 mL each) were collected and combined on the basis of TLC analysis, leading to five main series (A–E). Fractions 1–50, eluted with a mixture of hexane-EtOAc (9:1), gave series A, from which robustic acid (**2**) (4 g) crystallized. Fractions 101–120, eluted with hexane-EtOAc (3:2), gave series C, which was further subjected to repeated column chromatography over Si gel eluted with a mixture of hexane-EtOAc (7:3) to yield indicanine A (**1**) (70 mg). Series D, resulting from the combination of fractions 121–181 eluted with a mixture of hexane-EtOAc (1:1), was rechromatographed with Si gel column chromatography, eluting with hexane-EtOAc (3:2) to afford daidzein (60 mg) and 8-prenyldaidzein (100 mg).

**Indicanine A (1):** pale-yellow powder, mp 175–177 °C, yield 0.0012% [ $\alpha$ ]<sub>D</sub> –46° (c 1.99, MeOH); UV (MeOH)  $\lambda_{\text{max}}$  (log  $\epsilon$ ) 218 (4.54), 270 (3.48), 282 sh (3.86), 291 (4.23), 305 sh (4.17),

351 (4.63) nm; IR  $\nu_{\text{max}}$  (KBr) 3267, 1645, 1610, 1520, 1200, 1100 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz)  $\delta$  7.42 ppm (1H, d,  $J$  = 8.8 Hz, H-2' and H-6'), 6.93 (2H, d,  $J$  = 8.8 Hz, H-3' and H-5'), 6.58 (1H, s, H-8), 5.28 (1H, t,  $J$  = 8.8 Hz, H-2''), 5.0 (1H, d,  $J$  = 1.0 Hz, =CH), 4.95 (1H, d,  $J$  = 1.0 Hz, =CH), 4.08 (3H, s, 5-OMe), 3.80 (3H, s, 4'-OMe), 3.58 (1H, dd,  $J$  = 8.8 and 15.5 Hz, H-3''), 3.23 (1H, dd,  $J$  = 7.6 and 15.8 Hz, H-3''),  $\delta$  1.77 (3H, s, CH<sub>3</sub>-C=C); <sup>13</sup>C NMR (75 MHz)  $\delta$  164.1 (s, C-4), 162.7 (s, C-2), 161.1 (s, C-5), 158 (s, C-7), 155.1 (s, C-4'), 152.4 (s, C-8a), 142.4 (s, C-2''), 131.7 (d, C-2' and C-6'), 123.6 (s, C-1'), 113.5 (t, C-1''), 113.0 (d, C-3' and C-5'), 111.0 (s, C-6), 100.0 (s, C-5a), 86.1 (d, C-2''), 60.5 (q, 5-OMe), 55.2 (q, 4'-OMe), 33.2 (t, C-3'), 17.1 (q, 3''-Me); EIMS  $m/z$  [M]<sup>+</sup> 380 (96), 365 (33), 337 (19), 233 (100), 217 (33), 190 (44), 175 (28), 148 (98), 135 (33), 120 (41), 91 (30), 69 (43), 41 (21), 39 (16); DCI/NH<sub>3</sub> [M+1]<sup>+</sup> 381; HREIMS  $m/z$  [M<sup>+</sup>] 380.1262 (calcd for C<sub>22</sub>H<sub>20</sub>O<sub>6</sub>, 380.1264).

**Robustic acid (2):** white solid, yield 0.066%, mp 212° (lit.<sup>12</sup> 210°C); HEIMS  $m/z$  [M<sup>+</sup>] 380.1261 (calcd for C<sub>22</sub>H<sub>20</sub>O<sub>6</sub>, 380.1260); UV, IR, <sup>1</sup>H and <sup>13</sup>C NMR were in agreement with the published data.<sup>11,12</sup>

**Daidzein:** amorphous powder, yield 0.0001%, mp 198–200 °C; HREIMS  $m/z$  254.0577 (calcd for C<sub>15</sub>H<sub>10</sub>O<sub>4</sub>, 254.0579); IR, <sup>1</sup>H and <sup>13</sup>C NMR spectra data were in agreement with literature values.<sup>13</sup>

**8-Prenyldaidzein:** white amorphous solid, yield 0.0001%, mp 198 °C (lit.<sup>14</sup> 196–198 °C); HREIMS  $m/z$  322.1206 (calcd for C<sub>20</sub>H<sub>18</sub>O<sub>4</sub>, 322.1205); IR, <sup>1</sup>H and <sup>13</sup>C NMR spectral data matched well with those published in the literature.<sup>14</sup>

**Antimicrobial Activity Screening.** Extract and purified active compounds were tested at 1 mg/mL against *S. aureus* 209P, *M. smegmatis* ATCC 607, and *E. coli* RL65. The three strains of bacteria were cultured in Mueller–Hinton agar medium at 37 °C. After one day, their growth was assessed visually. The lowest concentration of the test compounds in which no visible growth occurred was defined as the minimum inhibitory concentration.

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## References and Notes

- Part 35 in the series "Erythrina Studies." For part 34, see: Nkengfack, A. E.; Vouffo, T. W.; Vardamides, J. C.; Kouam, J.; Fomum, Z. T.; Meyer, M.; Sterner, O. *Phytochemistry* **1997**, *46*, 573–578.
- Cordell, G. A. *Introduction to Alkaloids: A Biogenetic Approach*; John Wiley & Sons: New York, 1981; pp 450–462.
- Nkengfack, A. E.; Vardamides, J. C.; Fomum, Z. T.; Meyer, M. *Phytochemistry* **1995**, *40*, 1803–1808.
- Nkengfack, A. E.; Vouffo, W. T.; Fomum, Z. T.; Meyer, M.; Ola, B.; Sterner, O. *Phytochemistry*, **1994**, *36*, 1047–1051.
- Nkengfack, A. E.; Sanson, D. R.; Fomum, Z. T.; Tempesta, M. S. *J. Nat. Prod.* **1989**, *52*, 320–324.
- Fomum, Z. T.; Ayafor, J. F.; Mbafor, J. T.; Mbi, C. M. *J. Chem. Soc., Perkin Trans. 1* **1986**, 33–37.
- Mitscher, L. A.; Gollapudi, S. R.; Gerlach, D. C.; Drake, S. D.; Veliz, E. A.; Ward, J. A. *Phytochemistry* **1988**, *27*, 381–385.
- Kamat, V. S.; Chuo, F. Y.; Kubo, I.; Nakanishi, K. *Heterocycles* **1981**, *15*, 1163–1170.
- Ayenu, E. S. In *Medicinal Plants of West Africa*; Reference Publications, Inc.: Algonac, MI, 1978; p 153.
- Mitscher, L. A. In *Isolation, Separation and Purification of Antibiotics*; Weinstein, G., Wagman, G., Eds.; Elsevier: Amsterdam, 1977; p 463.
- Khalid, S. A.; Waterman, P. G. *Phytochemistry* **1983**, *22*, 1001–1003.
- Jackson, B.; Owen, P. J.; Scheinmann, F. *J. Chem. Soc. C* **1971**, 3389–3391.
- Jha, H. C.; Zillizen, F.; Breitmaier, V. *Can J. Chem.* **1980**, *58*, 1211–1213.
- Munekazu, I.; Toshiyuki, T.; Mizuo, M.; Hirobumi, Y.; Yasuko, K.; Shigetomo, Y. *Chem. Pharm. Bull.* **1997**, *40*, 2749–2752.
- Olivares, E. M.; Lwande, W.; Monache, F. D.; Bettolo, G. B. M. *Phytochemistry* **1982**, *21*, 1763–1765.
- East, A. J.; Ollis, W. D.; Wheeler, R. E. *J. Chem. Soc. C* **1969**, 365–367.

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